I'm not robot!

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Cuscuta Cuscuta is a stem holoparasitic plant without leaves or roots, which develops a haustorium and sucks nutrients from host plants. The genus Cuscuta begins in finding and attaching to a host plant and then developing a haustorium and sucks nutrients from host plants. The genus Cuscuta begins in finding and attaching to a host plant and then developing a haustorium and sucks nutrients from host plants.
The process does not always require any chemical signal, but does require a light signal. Finding a host involves detecting the lower red light ratio near a potential host plant by phytochrome. A contact signal is also necessary for haustorium induction. Apparently, cytokinin increase is downstream of the light and contact signal and is critical for
haustorium induction. This pathway, however, appears to be slightly different from a standard pathway. The direct connection between Cuscuta and its host involves both the xylem and phloem, and mRNA and proteins can translocate. Several features indicate that Cuscuta is a useful model plant for parasite plant research as well as plant¢ÃÂplant interaction
research. These include the simple anatomical structure and seedling development, no chemical requirement for haustorium induction, and the wide range of host plants. Begon et al. (2006) define parasites as organisms that obtain nutrients from a host and cause harm but not immediate death. The term ¢AAApathogen¢AAA may be applied to any parasite that
gives rise to a disease. Symbiotic relationships (living together), in contrast, refer to physical proximity; while mutualistic interaction means a conflict-free relationship. Some higher plants also have strategies that can be interpreted as being parasitic. These include myco-
 heterotrophic plants, twining plants, twining plants, and carnivorous plants. Myco-heterotrophic plants are typically considered to be mutualistic. Some have no leaves and Raãzes reduced (eg Burmannia Tenella) (Leake 1994). Superficially, certain morphological characteristics resemble other parasite plants. Myco-heterotrian plants were misunderstood as root parasites in their
 initial estate (Bidarndo 2005). Mico-heterotrian epiparasit plants appear to be developed from mutualist myo-heterotrian plants (Merckx et al. 2009). However, both Myco-Heterotrian plants groups absorb nutrients and water through fungi in host green plants without developing haustine. The specificity of the host is related to the generalist behavior of the shrub
mycorran fungi and is not always related to the self-heterotrian plants, but taxonhic relationships did not pose it (Heide-Jã £ £ 2008). Some parasitarian plants, including Custcuta string, to host plants, leading to comparison with intertwined/climbing plants. However,
the main strategy of delivery/climbing plants is effectively acquiring sunlight; They reduce sunlight to the plant, and this is not a mutualist. The intertwined/climbing plants are so leaves and bumps, but without haustian to obtain nutrients. Although certain © Cies of ivy develops staff similar, they are not able to suck nutrients (Heide-Jã £ £ 2008). Personal plants
have evolved independently at least six times and are specialized in insects. The acquisition of insect nutrients (Bauer and Federle 2009) resembles stratum from parasitarian plants, but with a great difference. Although the development of the throttle in nepents is energetically cheaper than the development of a normal green leaf, photosynthesis is strongly
correlated with carbon capture and of digested insects (ellison and gotelli 2009). Although the nutrients of insects are important for photosynthesis, these nutrients are insufficient for survival. on the other hand, many plants especially holoparasitic plants fully rely on host nutrients, and energy from photosynthesis appears to be secondary. Despite of some
similarity to other higher plants, parasitic plants have a unique strategy. The interaction between holoparasitic plants (e.g. Cuscuta) and host plants covers the full range from parasitic plants have a unique strategy. The interactions (Runyon et al. 2008), although the latter appear to be very rare. Recognition of parasitic plants itself was at a
quite early time. According to Heide-JÃ, Argensen (2008), Theophrastus (BC 372¢ÃÂÂ287) is the first person who described Cuscuta in Babylon, as a parasite plant. After the light microscope was introduced into botanic research mostly focused on the anatomy of the haustorium (Boewig 1898; Kindermann 1928). The
 haustorium of parasite plants is unique and posed an enigma as to how nutrients were absorbed from the host. The next step involved observing the host. AAAparasite plant connection. Hibberd and Jeschke (2001) stated that Solms-Laubach had already reviewed different types of parasite.
 1867. Despite the recognition of parasite plants at an early stage, studies tended to focus on each specific parasite plant, and no comprehensive study for all parasite plants was available until 1969. Biochemical approaches for parasite plant interactions are technically difficult, delaying research. Direct measurement of solute in xylem and
 phloem is complicated (Jeschke and Hilpert 1997), leading to other approaches such as those involving sap-sucking insects (Malone et al. 1967, 1968; Walting and Press 2001). More recently, such radiotracers have been replaced by xylem
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small in angiosperms () and the parasitic plants are evolved by the evolution of angiospermo. Cuscuta is classified in Family Convolvulaceae and the Ipomoeae tribe were relatively close to the couscutain gã. Anterior Molecular Philogan Research in sequences of internal nuclear ribs and plasticized sequences (for example, RPS2) (McNeal et al. 2007, 2009). The
couscuta gãa Nero consists of 3 subgathers, monogyna, couscuta and grammics (Mcneal et al. 2007), and between 150 and 200 spray have been described at now. The grammatic subgarten can be divided into eugrammic and clehistrophemics sections (Stefanovic et al. 2007). Download the CSVDiscaplay table in Principle, all forms of cuscuta develop a haustorium
which is a special fabric differentiated from the stem to suck nutrients from host plants. Although he has lost his leaves and grinds, couscuta is widespread in the world and commonly known as Dodder. This causes serious damage to certain cultures (eg tomatoes, potatoes, tobacco), especially in the US. There are several types of parasitic plants, for example and commonly known as Dodder. This causes serious damage to certain cultures (eg tomatoes, potatoes, tobacco), especially in the US.
 holoparasit or hemiparasitons, root parasites or STEM parasites or STEM parasites. The hemiparastic plants are defined as having chlorophyll and photosynthesis, holoparasitory plants as without these characteristics (Heide-Jã £ £ 2008). Stem parasites absorb nutrients and host rods, while the bumps make the host bumps. In general, the root/sheet can be reduced or lost in
parasitic plants, although this is different between the sprawles. For example, all couscuta and cassytha have not so leaves or bumps, but Olax still has functional bumps and many visitors to the leaves (Heide-Jã £ £ £ 2008). In Custuta, the green color in the stupid of seedlings indicates the preset of chlorophyll, but the generation is normally classified as
 holoparasit. Cuscuta seedlings usually live less than 3 weeks before they become parasitarian. The seedlings can not absorb the water (due to the lack of roots) and finally cannot survive without parasiticization. Besides, Cuscuta apparently exact before they become parasitarian. The seedlings can not absorb the water (due to the lack of roots) and finally cannot survive without parasiticization. Besides, Cuscuta apparently exact before they become parasiticization.
dnuof era snoitcennoc cimsalpmys dna cimsalpopA .tnalp tsoh eht rof noitcef suriv, and ,Birschwilks et al. (2006). The steps in the life cycle of parasitic plants include: (1) seed germination; (2) early development of the seed cultivated plant; (3) the search for a host plant, sewage induction and host invasion, sewage maturation; and (4) interaction with the host plant include: (1) seed germination; (2) early development of the seed cultivated plant; (3) the search for a host plant include: (1) seed germination; (2) early development of the seed cultivated plant include: (3) the search for a host plant include: (4) interaction with the host plant include: (5) early development of the seed cultivated plant include: (6) early development of the seed cultivated plant include: (7) early development of the seed cultivated plant include: (8) early development of the seed cultivated plant include: (9) early development of the seed cultivated plant include: (1) early development of the seed cultivated plant include: (1) early development of the seed cultivated plant include: (1) early development of the seed cultivated plant include: (1) early development of the seed cultivated plant include: (1) early development of the seed cultivated plant include: (1) early development of the seed cultivated plant include: (1) early development of the seed cultivated plant include: (2) early development of the seed cultivated plant include: (3) early development of the seed cultivated plant include: (4) early development of the seed cultivated plant include: (5) early development of the seed cultivated plant include: (6) early development of the seed cultivated plant include: (7) early development of the seed cultivated plant include: (8) early development of the seed cultivated plant include: (8) early development of the seed cultivated plant include: (8) early development of the seed cultivated plant include plant inc
 plant (Stewart and Press 1990; Yoder 1999). Various types of seed germination are known in parasitic plants (Press et al. 1990:) (1) Seeds are o covered with a fruit and eaten by birds. Consequently, the seeds are dispersed by birds and germinate in host plants (VISCUS,
 Santalaceae;) and (3) seed germination requires quantic products of the host plant root (orobanchaeceae) (Press et al. 1990). The size of the cscuuta seeds varies, but seem to contain enough nutrients. Early parasitic seedlings differ between spirit because hemiparasites can conduct photosynthesis, and may have leaves and grinds. In the case of couscuta, these of the cscuuta seeds varies, but seem to contain enough nutrients.
 initial seedlings are not a shape not embraced, similar to the rope; No leaf or root is present. Seeds grow up for vain days, so start to rotate to find a point of fixation. The activity of photosynthesis seems to be quite low due to a low affinity for CO2 (data not published). This makes it necessary to find a host plant within a few weeks. If the seedling is the change
cue important. Although this idea is attractive and widely accepted, it still needs more discussion. The authors said Custuta Campestris moved to host plants in the dark, due to the detection of voltable substances of host plants in the dark, due to the detection of voltable substances of host plants.
 random distribution (Figure 1). In fact, this rotation enables the seedlings to attach the plants hosted nearby. Another aspect is the variety of host plants for couscuta. Custuta can move from one host to another and go back. If the plant needs special -back products to look for a host, it is difficult to explain why it can parasitize so many different plants, except that
there is a strong overposure between the return compositions of the vain ¡Rias plants. In addition, the couscuta can do self-parasitization (Figure 2) and it can also move in direction to acril-free acril rod and induced cheamic products in vitro (Figure 3). This is an additional evidence that special chemicals are not necessarily necessary for the interaction of the
climbing plants can rotate even against the 'plastic materials, the chemicals may not be cructated for cscuuta (Figure 3). The word \phi and \phi and \phi and \phi and \phi are constant characteristics of parasite plants. This requires examining Haustorium's induction
 mechanism. Not hypothesized or data for the evolution of couscuta haustorium. Haustorium seems to have evolved a specialized fabric is not comparable to the meristem/side shot/lateral meristem are available about chemical cues for Cuscuta haustorium.
induction. The haustorium of Sriga is developed prior to attachment to the host. In Cuscuta the haustorium induced by light and a contact signal requires another chemical cue for maturation. One report
 describes that a haustorium can be induced on the apical part of Cuscuta without host attachment (Heide-JA, Argensen 2008), but this is based on field studies only. Cuscuta haustorium development progresses sequentially, and it continues even after detachment from a host plant or other object when sufficient stimuli are given (Tada et al. 1996). It is quite
commonly accepted that many higher plants avoid dense canopy conditions by detecting the change of red light: far-red light (R:FR) ratio by phytochrome (Pedmale et al. 2010). In fact, Cucumber (Cucumis sativus) de-ethiolated seedling experiments showed that seedlings bend away from a FR light source, which is comparable to a patchy canopy environment
 (BallarũÅ et al. 1992). Cuscuta conspicuously showed opposite behavior to a FR light source, and similar behavior as negative phototropism was reported only from some tropical vines, e.g. Monstera gigantean (Strong and Ray 1975). A photomorphogenic response, hook opening and circumnutating, and induction of twining by blue light and FR
light in Cuscuta were described in the past (Lane and Kasperbauer 1965) and later confirmed (Orr et al. 1996a, b). Lane and Kasperbauer already proposed a potential role of phytochrome in 1965 because of evidence that the R:FR ratio change in light is correlated with the phytochrome (e.g. phytochrome B) Pr and Pfr ratio. These studies, however, did not deal
with haustorium induction. Furuhashi et al. first reported in that that a light cue is necessary for Cuscuta japonica) haustorium induction than blue light only, although twining was almost the same
 induction as well as FR/red (R) light reversibility, clarifying that both the light signal and the contact signal are required for induction (Figure 4). Haidar et al. (1997) reported contradictory results for Cuscuta indecora and Cuscuta index in
2 min FR light did not induce a haustorium. Nevertheless, 1¢ÃÂ2 min of R or FR light treatment is no doubt too short, and the subsequent incubation and does not occur at longer irradiation. Indeed, Haidar et al. (1997) also showed
that longer irradiation (1¢ÃÂÂ2 days) with R light negatively affected haustorium induction but that FR light showed less induction but that Cuscuta seedlings treated solely with 100 µÂM m¢ÃÂÂ2 s¢ÃÂÂ1 of FR light, an
 intensity that may have been ineffective. The result showed that a high irradiance effect of FR light was not required for Cuscuta haustorium induction. The Pfr/Ptotal ratio is around 0.45 in plants growing muirotsuah o edseD. airotsuah revlovnesed mecerap atucsuC ed sadum, arugif aus me, ossid m©AlA. R zul ed otiefe on anAetorp ed o£A§Aartnecnoc ad
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 ronem ©Ã o£Ã§Ãroporp atse e ,luza zul It requires light and contact signal, an increase in protein concentration in blue light also partly reflecting the contact signal. A protein change related to a light sign is not conclusive and requires future studies. The density of light flow can also be a factor for parasitation. Custuts to develop much better haustority in
 obscure places than under strong direct sunlight (based on field observations and data on its sayings). This indicates that the sign of light for induction does not require a large density of light were sufficiently intensive, and it seems that couscuta prefers even weakens that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive, and it seems that couscuta prefers even weakens that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive, and it seems that couscuta prefers even weakens that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive, and it seems that couscuta prefers even weakens that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive, and it seems that couscuta prefers even weakens that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive, and it seems that couscuta prefers even weakens that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive, and it seems that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive, and it seems that suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive and suggested about 5-10 î¼m m-2 s-1 of blue or FR light were sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is sufficiently intensive and suggested about 5-10 if it is suffici
of light (light mixture R and FR in 200 î¼m m-2 Sâ € "1) In sewage induction was less than the lowest intensity (40 î¼m m-2 Sâ € "1). This indicates a greater light intensity of natural sunlight (approximately 2400 î¼m m-2 Sâ € "1) to shade (for example, 50% of full sunlight).
conditions (Clua et al. 2006). The percentage of each light is not always the same as blue (400 \stackrel{\epsilon}{} "700 nm), r (600 \stackrel{\epsilon}{} "700 nm), r (600 \stackrel{\epsilon}{} "700 nm), r (600 \stackrel{\epsilon}{} "700 nm) and the FR (700 \stackrel{\epsilon}{} "700 nm), r (600 \stackrel{\epsilon}{} "700 nm) and the FR (700 \stackrel{\epsilon}{} "700 nm) (Deitzer 1994). Relationship R: FR for the signal must be another important factor
because light r tends to inhibit, but light FR promotes the induction of the exhaust. Haidar and Orr (1999) reported a relationship R: FR between 0.02 and 1.0 )rohlem a iof Exhaust in Cuscuta planiflora. As Haidar and Orr (1999) pointed out, such conditions would exist in the proximity of other plants. In the case of Cuscuta, it is reasonable to develop
haustoria when and where the FR signal is increased. Thus, Cuscuta can detect the reflection of FR of nearby host plants in order to obtain more light, detecting a reduction in the R:FR ratio with phytochromia (Ballaré 2009). The shadow avoidance mechanism is probably similar in higher green
plants; Cuscuta may well have improved this system to recognize the proximity of potential hosts. In fact, the ratio R:FR both in the shadow and in the cups is less than 1, and the response to the shadow and sewage induction does not appear to be
 similar. The R:FR signal caused by the canopy tonality induces the production of auxin and gibberellin, but the induction of ehaustorium by Cuscuta seedlings did not significantly alter the concentrations of auxin (Löffler et al. 1999). The response to the canopy shade increases auxin and degrades cytokine, leading to the inhibition of radial growth and attenuation
of the development of leaves (Carabelli et al. 2008). In contrast, Haidar et al. (1998) reported that cytokine increases the development of haustorium of Cuscuta, but that auxin (as well as abscicus acid (ABA) and ethylene) inhibits it in blue light. Cuscuta has no roots, the site of cytokine production. This indicates that the cytokine level can be lower than in other
 higher green plants and can modify the hormonal signal transduction. In Cuscuta, the relationship between the FR signal and vegetable hormones implies that the eshaustorium would be completely different than in the meristem of the leaf or firing apical meristem. This, in turn, suggests that Cuscuta toohs/ahlof toohs/ahlof ad ocig³Älana uo golomoh selpmis mu
can influence at least two different signal transduction pathways (Monshausen and Gilroy 2009). One is the Ca2+> ion sensors (e.g. TCH and CML) interact with pinoids such as Ser/Thr kinase protein (PID). As the IAD can regulate the PIN family of auxin
regulators, auxin must be recognized downstream of mechanical detection and/or/o/o riopmomorphogenesis. Apparently, the concentration of auxin with the induction of Cuscuta Haustorium (Läffler et al. 1999). Another type of signal transduction
 depends on kinase. The cinases associated with the receiving type wall (WAKs) respond to mechanical stimulation, and this can be influenced by pectinestherosis and Ca2+>. Currently, we do not know what type is related to the induction of Cuscuta Haustorium. Overall, mechanical stress or stimulation negatively affects plants, inhibiting growth and decreasing
 pigments (e.g., anthocyanin). The fact that the Cuscuta stem connected to a host can change the color in pale is somewhat similar to previous studies. As cytokine can induce haustorium induction of the Haustorium induction signal and as being controlled by the signs to the color in pale is somewhat similar to previous studies. As cytokine can be predicted as the downstream of the Haustorium induction signal and as being controlled by the signs to the color in pale is somewhat similar to previous studies. As cytokine can induce haustorium induction signal and as being controlled by the signs to the color in pale is somewhat similar to previous studies.
 of light and contact (Figure 5). Otherwise, the increase in cytokine is spontaneous, but there is no substantial data to support this in natural conditions. As Cuscuta initially finds the host plant after detection of the FR light conditions, the light signal effect for induction can be prior to the contact signal in nature. After the induction of the Haustorium after the host
factor that activates these enzymes is present both in Cuscuta and in the host plant (Nun et al. 1999). Thus, the mechanism that helps Cuscuta avoid causing self-damage is still unknown (NUN and Mayer 1999). Pectinestherosis is present in bacteria and fungi and plays an important role for plant pathogens to penetrate or degrade the cell walls of the plant (e.g.
soft roots, vascular murches and foliage disease) (Wood 1960). The role of this enzyme appears to be quite similar in Cuscuta and bacteria/fungi as vegetable realm and is involved in seed germination, cellular adhesion and elongation of the stem with the increase of acidic pectin and decrease of
the cell wall. These can be influenced by vegetable hormones (e.g. auxin and giberellic acid) (Micheli 2001). Although cell-to-cell interactions or macromolecule (including viruses) through plasmodesmata are implicated with this enzymatic activity, substantial data is not available in relation to the plant interaction between Cuscuta and the host plant. Plant
host Xylem (Popp and Richter 1998). In xilema, the strong water current is one-way from the host to the parasite. On the other hand, a weaker current is bidirectional in the floema. Either way, Cuscuta connects her floema with the host to the parasite. On the other hand, a weaker current is bidirectional in the floema. Either way, Cuscuta connects her floema with the host to the parasite.
taken, without special selection (Figure 6). Plant¢ÃÂÂplant interactions between Cuscuta and host plants have recently been studied. These efforts revealed the translocation of mRNA and proteins between host and parasite. As outlined in a recent review, microarray experiments confirmed the presence of host plant mRNA in Cuscuta (Westwood et al. 2009)
Translocation of proteins from host (Arabidopsis) to Cuscuta has also been studied; the sieve size was predicted to be 27¢ÃÂÂ36 kDa based on GFP fused protein experiments (Birschwilks et al. 2007). However, plasmodesmata connections would not allow molecules over 800 Da to permeate without causing dilation (Haywood et al. 2002). Viral Movement protein
(MP) protein or non-cell-autonomous protein (NCAP) can induce microchannel dilation, but no information in this regard is known from Cuscuta is thought to mediate a flowering signal inhibitor from one host plant to another (Heide-JĀ Ârgensen 2008). As predicted, the sieve size in the phloem is 27¢ÂÂ36 kDa. Accordingly, one of the important
candidates of florigen named FT, which is expressed in phloem (Corbesier and Coupland 2006), among them sucrose, the plant hormone (e.g.
gibberellin, cytokinin, salicylic acid) pathways (Wada et al. 2010), and the circadian clock pathway. As salicylic acid is related to stress-induced flowering, there is a possibility that a pathological response of the host plant to Cuscuta might induce flowering. Unspecific movement of metabolites and macromolecules between parasite and host implies a problematic
relationship. One example involves parasite¢ÃÂĥost incompatibility. Incompatibility types can be classified into Mechanical defenses and defenses and defenses (e.g., formation or hypertrophy lignification). In an earlier study by Cuscuta, the antipathogenic test revealed that naftoquinone is one of the antipathogenic/antimicrobial substances produced by the host plant
(Ancistrocladus heyneanus). These substances trigger a subsequent hypersensitive reaction and lignification by the host plant. This was interpreted as a biochemical incompatibility for Cuscuta (Bringmann et al. 1999). These incompatible responses are similar to pathogenic responses. As such, some plant interactions resemble more pathogenic interactions than
symbiotics. Most pathogens involved in plant diseases involve signal transduction derived from eliciters (e.g. glucan). In the case of Cuscuta may be probable and cause incompatibility, but the data on this problem are rare. Ethylene inhibits cuscuta growth in the non-parasitic stage, but
the ethylene supplied exogenously has no effect in the parasitic phase (Haidar et al. 1998). This means that the increase in ethylene caused by the host's pathogenic response would not block Cuscuta's parasiticization. From the standpoint of the parasitic phase (Haidar et al. 1998). This means that the increase in ethylene caused by the host's pathogenic response would not block Cuscuta's parasitic phase (Haidar et al. 1998). This means that the increase in ethylene caused by the host's pathogenic response would not block Cuscuta's parasitic phase (Haidar et al. 1998).
apparently degrades the sorbitol absorbed by the host (Richter and Popp 1992; Wanek and Richter 1993). So far, no study suggests that Cuscuta degrades host metabolites or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolites or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolites or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolites or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolites or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolities or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolities or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolities or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolities or macromolecules, although the parasite probably has similar abilities to the visco. Cuscuta degrades host metabolities or macromolecules, although the parasite probably has similar abilities and although the parasite probably ha
germination. Its continuous growth and ability to successively change the hosts make it unlikely to occurAmong the cuscuta hosts and the specific hosts. No other mutant form without leaves and/or no threads have been reported between the highest plants. Only lateral root in oryza (wang et al. 2006) and loss of juvenile leaf in arabidopsis (hamada et al. 2000)
were reported, but this does not represent complete loss of u root leaf. this implies a singularity of the cuscut evolutionary strategy. also suggests difficulties in interpreting the system based only on mutant research involving model plants. no explanation for the loss of leaf and root in cuscuta was advanced, and no given fossil is available to suggest an
intermediate situation. So far, there is no clear idea how cuscuta evolved. the ancestral form can be a kind of gemination plant because cuscuta is philogenically near the ipomoea tribe. there is a report that ipomoea tribe and yellow,) indicating ipomoea can possibly oar light reflectance to look for
the structure to climb (price and wilcut 2007). photoreceptor (e.g. phytochrome) was also identified from ipomoea (lariguet and dunand 2005,) and it is conceivable that an ancestral form of cuscuta could have a similar host detection mechanism. a strong signal of contact by gemination for a host may have led to reduced chlorophyll, root and leaf development due
to mechanical stress. at this stage of evolution, certain individuals who could develop tissue as a hypha to suck nutrients would have been superior competitors. due to the fact that the sewage is a specialized tissue and not homologoo for leaves u roots, its evolution can occur parallel to the reduction u loss of the last two. Although light and contact signals are
necessary for exhaust induction, both the light signal and fr contact pathway to avoid the up-regulates auxin shade but cytokinin down-regulates. in cuscuta, however, the signal fr .)8991 .la te radiaH( oir3Åtsuahse on anicotic ad otnemua oa odived
anicotic a ratnemua The contact signal usually inhibits the growth of other superior plants, but increases the development of Cuscuta Haustorium. Therefore, the essential evolutionary change that is linked to the exclusive parasitic parasitic parasitic. A
direct connection with Floema may require the evolution of special enzymes to degrade metabs or macromolient unpleasant cules. Current model plant data can not be applied directly to the couscous parasitization mechanism and is stratum parasites. Other types of approaches are necessary to elucidate a wide range of biological fean. At the same time,
Cuscuta's research suggests that new aspects of light signal transduction and tigmomorphoga transduction in Custcuta. Some previous research on Cuscuta was based only on external observations. On the other hand, current molecular biological
research is based on specified conditions. These different research approaches are sometimes completely separated. In particular, plant molecular biology has only focused on some model plants (eg Arabidopsis). These models are suitable for understanding common biological characteristics, but not phenomena. In fact, no model plant is available that parasitizes
other plants. A current scientific question is how to use molecular biological data that is mainly obtained from model plants to understand other plants. This involves the comparison of customer research with model research. In addition, modern experimental data should be integrated into this venture, because our ultimate goal is also to explain phenomena
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 .icS geV locE .srorre gnippat-melyx fo ygoloisyhpocE .8991 .A, rethciR dna M .ppoP[ralohcS elgooG] .437-037 .81 .1018 rruC .sispodibarA fo stoor eht ni srosnes egatlov enarbmem amsalp .01LSMtA dna 9LSMtA dna 9
JY, sevarG, CM .108. 185. 482. [ralohcS elgooG] .858-748. (200. how in a setisarap mrepsoigna fo noitcaretni fo ygoloisyh. 200. (201. 185. 485. [ralohcS elgooG] .858-748. (201. 185. 486. [ralohcS elgooG] .858-748. (201. 186. [ralohcS elgooG]
:tsoh evitan dna decudortni fo seiceps a ni tnalp citisarap evitan a fo stcapmI .9002 MJ ,illecaF dna J ,goiltaW ,J ,redirP[ralohcS elgooG] .729-229 .(4)12 ,lonhceT deeW .stcejbo dna stnalp gnirobhgien ot (aecaredeh aeomopI) yrolggninrom faelyvi morf esnopseR .7002 WJ ,tucliW dna JA ,ecirP[ralohcS elgooG] .729-229 .(4)12 .[6 ecneicS fo beW] ,[ferssorC] .401-19 .[7 ecneicS fo beW] .tochec deed aeomopI) yrolggninrom faelyvi morf esnopseR .7002 WJ ,tucliW dna JA ,ecirP[ralohcS elgooG] .729-229 .[8]
ediug sekips lacimehc elitalov .6002 .DC ,searoM dna CM ,rehcseM ,BJ ,noynuR[ralohcS elgooG] .[871-184 .[121 .658. [203 .] ,noynuR[ralohcS elgooG] .[80, 204 .] ,retroin entiousive eht no siotilcyc fo noitalumucsive eht no siotilcyc fo noitalumucsive eht no siotilcyc fo noitalumuca eht fo ecnatropmi lacigoloisyhp eh .. (100 .] ,retroin ention elitalov .6002 .DC ,searoM dna CM ,rehcseM .[8] ,noynuR[ralohcS elgooG] .[80 .] ,retroin elitalov .6002 .DC ,searoM dna CM ,rehcseM .[8] ,noynuR[ralohcS elgooG] .[80 .] ,retroin ention entio
[deMbuP], [ferssorC] .599-789 :641 , loisyhP tnalp .serovibreh tcesni tsniaga stnalp elitsoh fo sesnefed eht setaunetta anogatnep atucsuC yb msitisarap ehT .8002 .DMC ,searoM dna CM ,rehcseM , [g. 1691-4691 :313 ,ecneicS fo beW], [deMbuP], [ferssorC] .599-789 :641 ,loisyhP tnalp .serovibreh tcesni tsniaga stnalp elitsoh fo sesnefed eht setaunetta anogatnep atucsuC yb msitisarap eht .8002 .DMC ,searoM dna CM ,rehcseM , [g. 1691-4691 :313 ,ecneicS fo beW]
oF infecoN by parasitiC angiospermS oN . 24â\(\epsilon\) 24â\(\epsilon\) 25., \(\tilde{a}\) 250., \(\tilde{a}\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoogle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoegle scholar[wanek, w and richteR, a. 1993. naD + 5-xidoreductase viscum album: utization oF host ., 5\(\phi\) algoegle scholar[wanek, w and richteR, algoegle scholar[wanek
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 [ralohcS [ralohcS elgooG]
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